

Spring 2013
Section 2387
Wednesday 2:30 – 4:25

The College of Staten Island
Department of Biology
Biology 107 – Principles of Biology I Laboratory

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REQUIRED TEXT: Laboratory Manual for Biology 10th or 11th Edition by Sylvia S. Mader
ISBN 978-0-07-747971-8 (11th ed.), ISBN 978-0-07-722617-6 (10th ed.)
Please bring the lab manual to each class period.

Course Description and Expectations

This course is designed to illustrate the principles taught in Bio 106. It is a laboratory course and not a lecture; thus, you are expected to read the material before coming to class so you can apply your knowledge to the lab activities. If you have questions on the reading, email me and I will be more than happy to help you. In addition to the required readings for each week, you may find it helpful to read the corresponding chapter in the 106 textbook.

During laboratory activities, you are expected to follow the given protocol in the lab manual and answer any questions given in the manual. Answering the questions in the lab manual will prepare you for both the quizzes and the final exam.

Grading

Your grade breakdown will be as follows:

Weekly Quizzes & Participation: 67%

Final Exam: 33%

Weekly Quizzes: COME PREPARED (do the required readings!) There will be a quiz at the start of lab every week lasting 15 minutes. If you miss the quiz period, you will receive a 0 for that quiz. The quizzes will be worth 10 points total – 5 points cover the reading assignment for the current week and 5 points reviewing the concepts from the previous week. This ensures that a) you are prepared for class and b) you understood the concepts learned the previous week. Showing up late will result in a missing quiz score. Quiz make-ups are not allowed without a doctor's note or previous arrangements. Please try to speak with me in advance about any conflicts.

Behavior and Participation Points: Your behavior and participation grade will be based on participation in the lab exercises and adherence to the rules. Points will be deducted for non-cooperative behavior. This includes leaving before completing the lab and failure to clean your lab bench. Distractions are not permitted. No use of cell phones, personal media players, food, or drink in the laboratory room. If you need a drink, snack, or bathroom break, you are free to do so outside of the room. Additionally, please see the academic honesty policy attached regarding cheating. Anyone caught cheating will be faced with disciplinary action.

Final Exam: A cumulative lab practical to be held on the last day of class.

Absence Policy

According to CUNY policy, more than two unexcused absences (15% of credit hours) **results in an automatic failure of the course.**

Biology 107 Lab Manual

Alan Tartakoff



Biology 107 Lab Manual:

Biology 107 Laboratory Manual for Texas A&M University Sylvia S. Mader, 2007 **Biology 107**, 2008 *Laboratory Methods in Vesicular and Vectorial Transport* Alan Tartakoff, 2012-12-02 *Laboratory Methods in Vesicular and Vectorial Transport* describes the procedures used to study the mechanisms of vesicular transport along the secretory and endocytic paths including electron microscopy autoradiography and methods associated with cyto and immunocytochemistry genetics and biochemistry It investigates vectorial transport to the cisternal space of the rough endoplasmic reticulum ER as well as protein translocation across the ER strategies for gaining access to the cytoplasm cell free analysis of vesicle fusion the structure of glycoproteins and the use of cell systems for analysis of vesicular traffic Organized into seven parts encompassing 20 chapters this volume begins with an overview of protein topology in the ER and the use of cross linking methods to probe the molecular environment of translocating polypeptide chains It then discusses the reconstitution of secretory protein translocation from detergent solubilized rough microsomes the use of anti idotype antibodies to characterize protein protein interactions the use of perforated cells to elucidate intracellular membrane transport delivery of macromolecules into cells expressing a viral membrane fusion protein and digitonin permeabilization procedures for studying endosome acidification and function The reader is also introduced to reconstitution of intracellular vesicle fusion in a cell free system after receptor mediated endocytosis immunoisolation using magnetic solid supports endosome and lysosome purification by free flow electrophoresis remodeling of glycoprotein oligosaccharides after endocytosis and replica plating of animal cells This book will interest students researchers geneticists biochemists and cell biologists The Buckeye, 1983

Antibodies in Cell Biology, 1993-11-17 *Antibodies in Cell Biology* focuses on a new generation of protocols aimed at the cell biologist This laboratory manual features systems and techniques that are especially relevant for modern problems The contributing authors have been carefully chosen for their specific expertise and have provided detailed protocols recipes and troubleshooting guides in each chapter The book is designed for any researcher or student who needs to use antibodies in cell biology and related research areas Practical applications and future emphases of antibodies including Light microscopic immunolocalization of antigens Gold particles in immunoelectron microscopy Special methods of fixation and permeabilization Microinjection of antibodies into living cells Antibodies to identify cDNA clones Antisense antibody strategies **Molecular Biology of the Cell**, 2003 **Bio 107 Laboratory Exercises** Montgomery College, 2003-08-18

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