

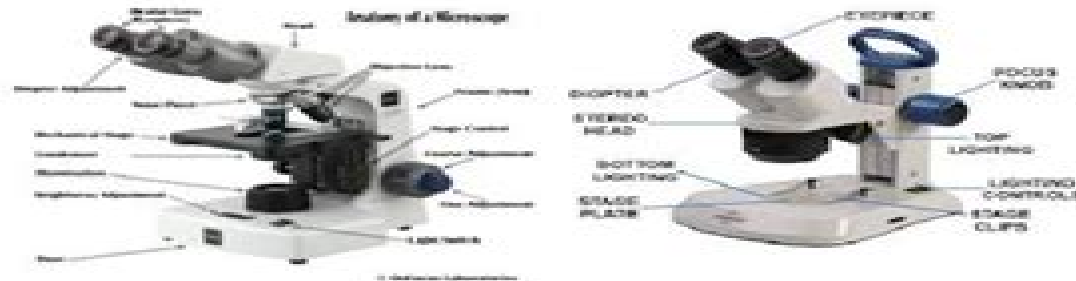
## BIO 108 Lab Exam Review

### 1. Define aseptic technique. How do you prepare an aseptic station and maintain aseptic technique?

Aseptic technique refers to a set of routine procedures done to prevent sterile solutions and cultures from becoming contaminated by unwanted microorganisms in the laboratory.

- decontaminate your lab bench
- safely organize your workspace
- properly adjust your Bunsen burner
- properly use an incinerator
- sterilize your inoculating tools
- aseptically transfer organisms from broth/plate cultures using procedures
- handle biohazard spills and dispose of biohazard materials
- do not put your head right over the things you are working on

### 2. Identify the parts of a compound and dissection microscope.



### 3. How do you calculate total magnification?

Ocular power x object power. Ocular power is 10x.

### 4. How do you calculate the actual size of an object? How do you complete the magnification of a drawing?

Actual size:  
 (1) actual measurement (with micrometer) or (2) objective field of view  
 Drawing size: 2000 μm (2 mm) → 1000 μm (1 mm) → 1000 μm (1 mm) → 1000 μm (1 mm)  
 Magnification:  
 Magnification =  $\frac{\text{Drawing Size}}{\text{Actual Size}}$   
 Magnification =  $\frac{1000 \mu\text{m}}{20 \mu\text{m}} = 50\times$   
 Magnification of object = 50x



### 5. How do you find the resolution of each objective lens?

| Objective Lens Magnification | Ocular Lens Magnification | Total magnification | Field of Diameter | Numerical Aperture (N.A.)<br><i>Focus on objective lens</i> | Resolution (R) | Calibration Factor<br>Ocular Micrometer Units (μm/ division) |
|------------------------------|---------------------------|---------------------|-------------------|-------------------------------------------------------------|----------------|--------------------------------------------------------------|
| 40x                          | 10x                       | 400x                | 3.6mm             | 0.10                                                        | 3.6 μm         | 1 ocular = 30.8 μm                                           |
| 100x                         | 10x                       | 1000x               | 1.8mm             | 0.25                                                        | 1.0 μm         | 1 ocular = 10.2 μm                                           |
| 400x                         | 10x                       | 4000x               | 0.45mm            | 0.65                                                        | 0.4 μm         | 1 ocular = 2.5 μm                                            |
| 1000x                        | 10x                       | 10000x              | 0.18mm            | 1.25                                                        | 0.2 μm         | 1 ocular = 1.0 μm                                            |

$$R = \frac{0.61 \lambda}{N.A.} \quad \text{Where } \lambda = \text{wavelength of light (500 nm or } 0.5 \mu\text{m)}$$

# Bio 108 Lab Manual

**Siddappa N.Byraredddy**



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liberation of proteins from cellular material removal of lipids from lipoproteins and denaturation fractionation and purification of proteins The next chapter focuses on the isolation of biologically active proteins such as cytochrome bacterial amylases and bacterial proteinases The reader is methodically introduced to fractionation of proteins by adsorption and ion exchange fractionation and characterization by dialysis multi membrane electrodecantation and continuous and discontinuous partition The final chapter explains how zonal density gradient electrophoresis works as a separation method for natural mixtures of proteins their degradation products and other substances carrying electric charges in solution or suspension This volume will be of interest to chemists working with proteins

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