

MJPRU BAREILLY

BSc 3rd & 5th sem Syllabus

BSc 3rd & 5th sem Zoology Syllabus

5th सेमेस्टर में 3 पेपर होंगे 👍

जल्दी देखो

क्या-क्या बदल गया 👍

Programme/Course Degree	Year: Third	Semester: Fifth
Subject: ZOOL001		
Course Code: B0405021	Course Title: Diversity of Chordates and Comparative Anatomy	
Course objectives: A student at the completion of the course will be able to:		
• Demonstrate comprehensive identification abilities of chordate diversity • Explain structural and functional diversity of chordates • Explain evolutionary relationship amongst chordates • Take up research in biological sciences.		
Credits: 4		Core/Compulsory/Elective
Max. Marks: 25x5		Min. Passing Marks: as per rules
Total No. of Lectures/Tutorials/Practical (in hours per week) : L/T/P: 4/0/0		
Unit	Topic	Total No. of Lectures (Hr)
1	Origin of Chordates & Hemichordates • Origin of Chordates: Classification of Phylum Chordata upto the class. • Hemichordates: General characteristics, classification and detailed study of <i>Balanophthalmus</i> and <i>Holothur</i>. (Morphology, Anatomy, Physiology, and Development).	4
2	Cephalochordates and Cephalochordates • Cephalochordates: General characteristics, classification and detailed study of <i>Balanophthalmus</i> (<i>Amphioxus</i>) (Habit and Habitat, Morphology, Anatomy, Physiology). • Cephalochordates: General characteristics, classification and detailed study of <i>Amphioxus</i> (Habit and Habitat, Morphology, Anatomy, Physiology).	4

Bsc 3rd Year 5th Sem Lab Manual

N Colangelo



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Instrumentation Debasish Mondal, 2024-05-04 Here s a brief description of each unit Unit 1 Microscopy Brightfield and darkfield microscopy Basic techniques for observing biological samples with and without staining Fluorescence Microscopy Visualization of fluorescently labeled molecules in biological samples Phase contrast Microscopy Enhancing contrast in transparent specimens Confocal Microscopy High resolution imaging technique with optical sectioning capability Electron Microscopy Scanning and Transmission Electron Microscopy High resolution imaging using electron beams Micrometry Measurement of microscopic objects and structures Unit 2 Chromatography Principles and applications of various chromatographic techniques Paper chromatography Thin layer chromatography Column packing and fraction collection Preparation and separation of compounds in columns Gel filtration chromatography Separation based on molecular size Ion exchange chromatography and affinity chromatography Separation based on charge and specific interactions Gas liquid chromatography GLC and High performance liquid chromatography HPLC Separation based on different principles Unit 3 Electrophoresis Principles and applications of various electrophoretic techniques Polyacrylamide gel electrophoresis SDS polyacrylamide gel electrophoresis 2D gel electrophoresis Isoelectric focusing Separation based on differences in isoelectric points Zymogram preparation Detection of enzymatic activity in electrophoresis gels Agarose gel electrophoresis Separation of nucleic acids based on size Unit 4 Spectrophotometry Principles of absorption spectroscopy Measurement of light absorption by biomolecules UV and visible range analysis Quantification of biomolecules based on absorption in UV and visible spectra Colorimetry and turbidometry Measurement of color changes and turbidity in biochemical assays Unit 5 Centrifugation Preparative and analytical centrifugation Separation of particles based on density and size Fixed angle and swinging bucket rotors Different configurations for centrifugation RCF Relative Centrifugal Force and sedimentation coefficient Parameters used to characterize centrifugation Differential centrifugation and density gradient centrifugation Techniques for separating particles based on density Ultracentrifugation High speed centrifugation for studying biomolecules and subcellular components

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